

# Biological Pretreatment Strategies For Enhanced Saccharification Of Lignocellulosic Biomass In 2G Ethanol Biorefineries

Diksha Sharma<sup>1</sup>, Rohit Rai<sup>2</sup>

<sup>1</sup>Department of Biotechnology, DAV College Jalandhar, India-144008

<sup>2</sup>Faculty of Applied Medical Sciences, Lovely Professional University, Phagwara, India-144411

Email : rohitraisharma44@gmail.com

## ABSTRACT

*Rapidly increasing demand and depleting reserves of crude oil have pushed scientific community to work towards finding alternatives of the fossil fuels. Lignocellulosics based 2G ethanol is being looked as a sustainable eco-friendly alternative of the crude oil to be utilized in transportation sector. Despite decade-ful of research on developing 2G biorefineries, pretreatment of LCB still remains one of the major bottlenecks. There are several physical and chemical pretreatment methods in practice since years but high process cost, production of fermentation inhibitors and toxic waste generation are some of the major concerns associated with them. Biological pretreatment is a good alternative in this direction which offers a greener and cleaner pathway to get rid of recalcitrant lignin fraction that obstructs the access of cellulolytic enzymes to target sites present in LCB. Although biological pretreatment strategies are being explored from past couple of decades but development of an economically viable and efficient technique is still under research. The recent developments in this field have indicated towards formulation of bacterial co-cultures, fungal co-cultures and bacteria-fungi co-cultures to attain efficient bioconversion of lignocellulosic biomass into ethanol. Through this article, an effort is made to review various biological pretreatment strategies in practice with main emphasis on the enzyme and microorganisms involved, regulation of ligninolytic enzymes, and process parameters affecting the success of the strategy adopted.*

**Keywords:** *Biological pretreatment, saccharification, lignocellulosic biomass, regulation, consortium, co-culture, laccase, lignin*

## 1. INTRODUCTION

Pretreatment of lignocellulosic biomass (LCB) is one of the three essential steps: pretreatment, hydrolysis and fermentation, involved in the production of 2G biofuels (Xiao, Yin, Xia, & Ma, 2012). The breakdown of LCB into fermentable sugars is restricted strongly by its crystallinity, degree of recalcitrance and polymerization with lignin being the major hindering moiety (Kim & Lee, 2006; Yang & Wyman, 2006). Lignin limits the second step of biorefineries i.e., hydrolysis by binding in a non-productive manner to the hydrolytic enzymes (Esteghlalian, Hashimoto, Fenske, & Penner, 1997). Numerous pretreatment strategies such as grinding, milling, chopping, liquid hot water treatment, ammonia fiber explosion (AFEX), dilute ammonia, ionic liquids, organosolv process, alkali, acid and biological treatment have been used by the researchers to disrupt interactions of lignin with carbohydrate fraction for overall improved hydrolysis yield (Alvira, Tomás-Pejó, Ballesteros, & Negro, 2010; Keshwani, 2009; Socha et al., 2014; Taherzadeh & Karimi, 2008). Among all the above stated strategies, biological pretreatment of LCB seems to be a potent mild and

eco-friendly method that does not involve generation of fermentation inhibitors and toxic waste products. Moreover, biological pretreatment strategies involve lesser cost and energy inputs (Sharma, Xu, & Qin, 2019). The biological pretreatment strategies rely mostly on fungi and bacteria which can efficiently degrade lignin, hemicellulose and little of the cellulosic fraction (Sánchez, 2009). The studies have established white rot fungi as the most efficient biological technique to delignify LCB through the action of lignin peroxidases, manganese peroxidases, and laccases (Kumar & Wyman, 2009; Shi, Chinn, & Sharma-Shivappa, 2008). Recent studies also indicate towards the potential of bacterial enzyme systems to efficiently treat LCB for enhanced sugar yields (Verma & Shirkot, 2014). The concept of consortia and co-culture is also into practice by many researchers which not only bring the diverse catalytic machineries on a common platform, helps in enhancing the lignocellulosytic abilities of different microorganisms as well. The platform developed therefore improves the yield of monomeric sugars resulting in improved subsequent 2G ethanol yield. However, long residence time for effective delignification and ability of the microbes to attack cellulose and hemicellulose fractions of biomass are couple of limitations associated with the biological pretreatment strategies. These biological strategies are not fully exploited yet, therefore, there is need for an extensive research in this field to develop an economically viable process. This review summarizes various biological treatment methods addressing the responsible microorganisms, associated enzymes and their regulation.

## 2. BIOLOGICAL PRETREATMENT OF LIGNOCELLULOSIC BIOMASS

Pretreatment of LCB is first and the most important step of 2G biofuel production since it constitutes nearly 40% of the overall process cost (Zhang, Li, Shen, Wang, & Sun, 2000). The importance of pretreatment step can be judged from the fact that yield of fermentable sugars post treatment increases upto 70% as compared to untreated biomass (Alizadeh, Teymouri, Gilbert, & Dale, 2005). The fermentable cellulose fraction in LCB is naturally protected by hemicellulose and lignin which reduces surface area accessible to cellulolytic enzymes. Therefore, proper pretreatment of LCB is necessary to increase the concentration of monomeric sugars which can be subsequently fermented into ethanol. The main enzymes involved in lignin digestion are peroxidases and laccases where former has lignin peroxidase and manganese peroxidase enzymes capable of degrading non-phenolic and phenolic lignin units, respectively. Laccases on the other hand act synergistically with peroxidases to degrade lignin completely. However, recent studies have shown that laccases can single handedly take care of the lignin fractions of LCB (Binod, Janu, Sindhu, & Pandey, 2011). The auxiliary activity (AA) proteins including lytic polysaccharide monooxygenases (LPMOs) and cellobiose dehydrogenases (CDHs) have been reported to increase the enzymatic hydrolysis of different lignocellulosic substrates (Levasseur, Drula, Lombard, Coutinho, & Henrissat, 2013). The bacterial and fungal strains known to hydrolyze recalcitrant biopolymers are discussed in the following sections.

### 2.1. Bacterial strains

There are several bacteria reported for their biomass degrading potential but screening of best bacterial strain to be employed for pretreatment of LCB in the biorefineries is the most critical step for 2G ethanol production. For over several decades' fungal lignin degrading enzymes were in trend but recently the paradigm has shifted towards bacterial enzyme systems owing to their better thermal stability. The cellulolytic bacteria *Cellulomonas fimi*, *Thermomonosporafusca* and *Paenibacillus campinasensis* have been showcased in past for their potential to pretreat LCB (Maki, Leung, & Qin, 2009; Sharma et al., 2019). Bacterial species such as *F. succinogenes*, *R. albus*, *R. flavefaciens*, etc. are associated with the rumen and show enormous potential to adhere with cellulose and mediate its hydrolysis (Duff &

Murray, 1996). Some of the bacteria like *A. lipoferum* and *B. subtilis* have shown delignifying abilities because of the laccases they produce (Saritha & Arora, 2012).

### 2.2. Fungal strains

Fungi are the intensively explored microorganisms for their potential to produce ligninolytic enzymes. Most of the lignocellulolytic fungi belong to genus *Aspergillus*, *Penicillium*, *Trichoderma*, *Schizophyllum*, *Fomitopsis*, *Orpinomyces*, *Trametes*, etc., (Dashtban, Schraft, & Qin, 2009; Paudel & Qin, 2015; Rai, Kaur, Singh, et al., 2016). Degradation of lignin on the other hand is a complex process and its success depends largely upon the selection of fungal strain. White rot fungi with special reference to basidiomycetes have significant lignin disintegrating powers and are considered as natural degraders of lignin. A study revealing 30 wood decaying white rot fungi was conducted in which *Phellinus pni-2*, *Pholiotamutabilis*, *Phlebia brevispora-1*, *Phanerochaete chrysosporium* were reported as the best delignifiers (Otjen, Blanchette, Effland, & Leatham, 1987).

### 2.3. Other macroorganisms

Apart from microorganisms there are several macroorganisms which have enormous potential to degrade lignocelluloses. These organisms include insects, worms, gastropods and ruminant animals which possess different masticating mechanisms for physical breakdown of recalcitrant biomass and enzymatic machinery for digestion of cellulose. There are nearly 20 families of insects including beetles, termites, wasps, silverfish, cricket etc., that can degrade leaf litters, forage and wood (J. Sun, Ding, & Doran-Peterson, 2013). The enzymatic activities within the gut of earthworms such as *Perionyx excavates*, *Lumbricus rubellus*, *Eisenia fetida*, etc., are known to mediate digestion of cellulose, chitin, lignin, starch, sugars, etc., (Pathma & Sakthivel, 2012; Vivas, Moreno, Garcia-Rodriguez, & Benítez, 2009; Zhang et al., 2000). Worm tea, the liquid leachate of vermicomposting is considered as a microbial consortium and has been used as an alternative of acid pretreatment for biofuel production (Siti, Siti, Nur, Renuka, & Norli, 2013). On the similar grounds, microbial consortia of gastropods and ruminants also have the potential to degrade lignin fraction of LCB (Fondevila & Dehority, 1994; Russell, Muck, & Weimer, 2009; Weimer, Nerdahl, & Brandl, 2015).

### 2.4. Microbial co-cultures

In biological pretreatment strategies, high enzyme activity is always desired, however, it is not always possible to obtain such higher titers of lignocellulolytic enzymes from a single bacterial or fungal strain. Therefore, concept of co-culture of two or more microbial strains can be very useful in achieving significantly higher activities of all the desired enzymes. There could be three possible combinations of bacterial and fungal strains for performing pretreatment of LCB.

Bacterial co-cultures involving two or more bacterial species can be useful in biofuel production. Many bacterial genera such as *Cellulomonas*, *Clostridium*, *Thermomonospora*, *Bacillus*, *Streptomyces* and *Ruminococcus* produce various cellulases that can work together to carry out cellulolytic hydrolysis (Y. Sun & Cheng, 2002; Zhou & Ingram, 2000). A study showed significantly higher hydrolytic potential when *Paenibacillus* sp., *Bacillus* sp., and *Aneurinibacillus aneuriniticus* were cultured together in comparison to their pure cultures (Chandra & Chowdhary, 2015). The study on co-cultures of *Clostridium thermocellum* and other *Clostridia* has exhibited significant increase in cellulolytic/hemicellulolytic hydrolysis into fermentable sugars (Maki et al., 2009).

Fungal co-culture has been in practice for pretreatment of LCB from past couple of decades. A study on co-culture of *T. reesei* and *A. phoenicis* exhibited high levels of total cellulase and  $\beta$ -glucosidase activities whereas the monoculture of two fungi showed high total cellulase and low  $\beta$ -glucosidase activity, and low total cellulase and high  $\beta$ -glucosidase activity, respectively (Wen, Liao, & Chen, 2005). Further, a study showed that lignin

degradation improved significantly in a co-culture of *C. subvermispora* and *P. ostreatus* when compared their monocultures (Chi, Hatakka, & Majjala, 2007).

There is a recent shift in the paradigm from bacterial or fungal co-cultures to co-cultures involving both bacterial and fungal strains. This type of co-culture mimics nature where different microbial communities co-exist and communicate via interconnected networks to derive their nourishment, breaking down complex substrates into simpler ones. Examples of such techniques can be found in literature where *T. reesei* and *E. coli* were co-cultured to produce isobutanol and *Z. mobilis* and *P. stipites* were co-cultured to produce ethanol (Fu, Peiris, Markham, & Bavor, 2009; Minty et al., 2013)

### **3. REGULATION OF ENZYMES INVOLVED IN BIOLOGICAL PRETREATMENT**

Lignin degradation potential of microorganisms can be improved significantly by employing molecular techniques. The regulatory elements present in promoter region play an important role in regulating expression of ligninolytic enzymes. The expression of *P. chrysosporium* genes has been reported to be positively influenced through carbon and nitrogen limitation (Cohen, Hadar, & Yarden, 2001). In a protein expression study, researchers have shown the effect of different carbon sources on the expression of lignocellulolytic enzymes (Rai, Kaur, & Chadha, 2016). The effect of different substrates on the expression of genes involved in lignin degradation has also been reported in literature (Salame, Yarden, & Hadar, 2010).

### **4. FACTORS AFFECTING BIOLOGICAL PRETREATMENT**

Although biological pretreatment strategies are greener and cleaner approaches which do not generate any fermentation inhibitor, long treatment time involved is a major limiting factor. However, screening of the most efficient microbial strains and optimization of the culture conditions can increase efficiency of the overall process. Apart from the composition of LCB, several process parameters like treatment temperature, pH, incubation time, inoculum age and concentration, moisture content and rate of aeration are the important factors governing the success of designed biological pretreatment strategy (Du et al., 2011; Isroi et al., 2011; Patel, Gupte, & Gupte, 2009).

### **5. CONCLUSIONS AND FUTURE PERSPECTIVES**

Lignocellulosics based (2G) ethanol is being sorted as an alternative source of renewable energy, however, development of an economically viable and efficient pretreatment technology is still needed. Since biological methods have several advantages over the other physical and chemical methods of pretreatment, therefore, addressing challenges associated with the former to reduce cost and time should be the main focus of future research.

### **6. ACKNOWLEDGEMENT**

The authors did not receive any external funding for this work. We acknowledge the contribution of each author.

#### *Conflict of interest*

The authors declare no conflict of interest.

### **7. REFERENCES**

- [1] Alizadeh, H., Teymouri, F., Gilbert, T. I., & Dale, B. E. (2005). Pretreatment of switchgrass by ammonia fiber explosion (AFEX). *Applied Biochemistry and Biotechnology*, 124(1-3), 1133-1141.

- [2] Alvira, P., Tomás-Pejó, E., Ballesteros, M., & Negro, M. (2010). Pretreatment technologies for an efficient bioethanol production process based on enzymatic hydrolysis: a review. *Bioresource technology*, 101(13), 4851-4861.
- [3] Binod, P., Janu, K., Sindhu, R., & Pandey, A. (2011). Hydrolysis of lignocellulosic biomass for bioethanol production *Biofuels* (pp. 229-250): Elsevier.
- [4] Chandra, R., & Chowdhary, P. (2015). Properties of bacterial laccases and their application in bioremediation of industrial wastes. *Environmental Science: Processes & Impacts*, 17(2), 326-342.
- [5] Chi, Y., Hatakka, A., & Maijala, P. (2007). Can co-culturing of two white-rot fungi increase lignin degradation and the production of lignin-degrading enzymes? *International Biodeterioration & Biodegradation*, 59(1), 32-39.
- [6] Cohen, R., Hadar, Y., & Yarden, O. (2001). Transcript and activity levels of different *Pleurotus ostreatus* peroxidases are differentially affected by Mn<sup>2+</sup>. *Environmental microbiology*, 3(5), 312-322.
- [7] Dashtban, M., Schraft, H., & Qin, W. (2009). Fungal bioconversion of lignocellulosic residues; opportunities & perspectives. *International journal of biological sciences*, 5(6), 578.
- [8] Du, W., Yu, H., Song, L., Zhang, J., Weng, C., Ma, F., & Zhang, X. (2011). The promoting effect of byproducts from *Irpex lacteus* on subsequent enzymatic hydrolysis of bio-pretreated cornstalks. *Biotechnology for biofuels*, 4(1), 37.
- [9] Duff, S. J., & Murray, W. D. (1996). Bioconversion of forest products industry waste cellulose to fuel ethanol: a review. *Bioresource technology*, 55(1), 1-33.
- [10] Esteghlalian, A., Hashimoto, A. G., Fenske, J. J., & Penner, M. H. (1997). Modeling and optimization of the dilute-sulfuric-acid pretreatment of corn stover, poplar and switchgrass. *Bioresource technology*, 59(2-3), 129-136.
- [11] Fondevila, M., & Dehority, B. (1994). Degradation and utilization of forage hemicellulose by rumen bacteria, singly in coculture or added sequentially. *Journal of applied bacteriology*, 77(5), 541-548.
- [12] Fu, N., Peiris, P., Markham, J., & Bavor, J. (2009). A novel co-culture process with *Zymomonas mobilis* and *Pichia stipitis* for efficient ethanol production on glucose/xylose mixtures. *Enzyme and microbial technology*, 45(3), 210-217.
- [13] Isroi, I., Millati, R., Niklasson, C., Cayanto, C., Taherzadeh, M. J., & Lundquist, K. (2011). Biological treatment of Lignocelluloses with white-rot fungi and its applications. *BioResources*, 6(4), 5224-5259.
- [14] Keshwani, D. R. (2009). Microwave pretreatment of switchgrass for bioethanol production.
- [15] Kim, T. H., & Lee, Y. (2006). Fractionation of corn stover by hot-water and aqueous ammonia treatment. *Bioresource technology*, 97(2), 224-232.
- [16] Kumar, R., & Wyman, C. E. (2009). Effects of cellulase and xylanase enzymes on the deconstruction of solids from pretreatment of poplar by leading technologies. *Biotechnology progress*, 25(2), 302-314.
- [17] Levasseur, A., Drula, E., Lombard, V., Coutinho, P. M., & Henrissat, B. (2013). Expansion of the enzymatic repertoire of the CAZy database to integrate auxiliary redox enzymes. *Biotechnology for biofuels*, 6(1), 41.
- [18] Maki, M., Leung, K. T., & Qin, W. (2009). The prospects of cellulase-producing bacteria for the bioconversion of lignocellulosic biomass. *International journal of biological sciences*, 5(5), 500.
- [19] Minty, J. J., Singer, M. E., Scholz, S. A., Bae, C.-H., Ahn, J.-H., Foster, C. E., . . . Lin, X. N. (2013). Design and characterization of synthetic fungal-bacterial consortia

for direct production of isobutanol from cellulosic biomass. *Proceedings of the National Academy of Sciences*, 110(36), 14592-14597.

- [20] Otjen, L., Blanchette, R., Effland, M., & Leatham, G. (1987). Assessment of 30 white rot basidiomycetes for selective lignin degradation: Walter de Gruyter, Berlin/New York.
- [21] Patel, H., Gupte, A., & Gupte, S. (2009). Effect of different culture conditions and inducers on production of laccase by a basidiomycete fungal isolate *Pleurotus ostreatus* HP-1 under solid state fermentation. *BioResources*, 4(1), 268-284.
- [22] Pathma, J., & Sakthivel, N. (2012). Microbial diversity of vermicompost bacteria that exhibit useful agricultural traits and waste management potential. *SpringerPlus*, 1(1), 26.
- [23] Paudel, Y., & Qin, W. (2015). Two bacillus species isolated from rotting wood samples are good candidates for the production of bioethanol using agave biomass. *Journal of Microbial and Biochemical Technology*, 7(4), 218-225.
- [24] Rai, R., Kaur, B., & Chadha, B. (2016). A method for rapid purification and evaluation of catalytically distinct lignocellulolytic glycosyl hydrolases from thermotolerant fungus *Acrophialophora* sp. *Renewable Energy*, 98, 254-263.
- [25] Rai, R., Kaur, B., Singh, S., Di Falco, M., Tsang, A., & Chadha, B. (2016). Evaluation of secretome of highly efficient lignocellulolytic *Penicillium* sp. Dal 5 isolated from rhizosphere of conifers. *Bioresource technology*, 216, 958-967.
- [26] Russell, J. B., Muck, R. E., & Weimer, P. J. (2009). Quantitative analysis of cellulose degradation and growth of cellulolytic bacteria in the rumen. *FEMS Microbiology Ecology*, 67(2), 183-197.
- [27] Salame, T. M., Yarden, O., & Hadar, Y. (2010). *Pleurotus ostreatus* manganese-dependent peroxidase silencing impairs decolourization of Orange II. *Microbial biotechnology*, 3(1), 93-106.
- [28] Sánchez, C. (2009). Lignocellulosic residues: biodegradation and bioconversion by fungi. *Biotechnology advances*, 27(2), 185-194.
- [29] Saritha, M., & Arora, A. (2012). Biological pretreatment of lignocellulosic substrates for enhanced delignification and enzymatic digestibility. *Indian journal of microbiology*, 52(2), 122-130.
- [30] Sharma, H. K., Xu, C., & Qin, W. (2019). Biological pretreatment of lignocellulosic biomass for biofuels and bioproducts: an overview. *Waste and Biomass Valorization*, 10(2), 235-251.
- [31] Shi, J., Chinn, M. S., & Sharma-Shivappa, R. R. (2008). Microbial pretreatment of cotton stalks by solid state cultivation of *Phanerochaete chrysosporium*. *Bioresource technology*, 99(14), 6556-6564.
- [32] Siti, N., Siti, A., Nur, F., Renuka, R., & Norli, I. (2013). Second generation bioethanol from lignocellulosic biomass using worm tea as pretreatment. *International Proceedings of Chemical, Biological and Environmental Engineering (IPCBE)*, 58, 1-5.
- [33] Socha, A. M., Parthasarathi, R., Shi, J., Pattathil, S., Whyte, D., Bergeron, M., . . . Venkatachalam, S. (2014). Efficient biomass pretreatment using ionic liquids derived from lignin and hemicellulose. *Proceedings of the National Academy of Sciences*, 111(35), E3587-E3595.
- [34] Sun, J., Ding, S.-Y., & Doran-Peterson, J. (2013). Biomass and its biorefinery: novel approaches from nature-inspired strategies and technology *Biological Conversion of Biomass for Fuels and Chemicals* (pp. 1-13).
- [35] Sun, Y., & Cheng, J. (2002). Hydrolysis of lignocellulosic materials for ethanol production: a review. *Bioresource technology*, 83(1), 1-11.

- [36] Taherzadeh, M. J., & Karimi, K. (2008). Pretreatment of lignocellulosic wastes to improve ethanol and biogas production: a review. *International journal of molecular sciences*, 9(9), 1621-1651.
- [37] Verma, A., & Shirkot, P. (2014). Purification and Characterization of Thermostable Laccase from Thermophilic *Geobacillus thermocatenulatus* MS5 and its applications in removal of Textile Dyes. *Sch Acad J Biosci*, 2(8), 479-485.
- [38] Vivas, A., Moreno, B., Garcia-Rodriguez, S., & Benítez, E. (2009). Assessing the impact of composting and vermicomposting on bacterial community size and structure, and microbial functional diversity of an olive-mill waste. *Bioresource technology*, 100(3), 1319-1326.
- [39] Weimer, P. J., Nerdahl, M., & Brandl, D. J. (2015). Production of medium-chain volatile fatty acids by mixed ruminal microorganisms is enhanced by ethanol in co-culture with *Clostridium kluyveri*. *Bioresource technology*, 175, 97-101.
- [40] Wen, Z., Liao, W., & Chen, S. (2005). Production of cellulase/ $\beta$ -glucosidase by the mixed fungi culture *Trichoderma reesei* and *Aspergillus phoenicis* on dairy manure. *Process Biochemistry*, 40(9), 3087-3094.
- [41] Xiao, W., Yin, W., Xia, S., & Ma, P. (2012). The study of factors affecting the enzymatic hydrolysis of cellulose after ionic liquid pretreatment. *Carbohydrate Polymers*, 87(3), 2019-2023.
- [42] Yang, B., & Wyman, C. E. (2006). BSA treatment to enhance enzymatic hydrolysis of cellulose in lignin containing substrates. *Biotechnology and Bioengineering*, 94(4), 611-617.
- [43] Zhang, B.-G., Li, G.-T., Shen, T.-S., Wang, J.-K., & Sun, Z. (2000). Changes in microbial biomass C, N, and P and enzyme activities in soil incubated with the earthworms *Metaphire guillelmi* or *Eisenia fetida*. *Soil Biology and Biochemistry*, 32(14), 2055-2062.
- [44] Zhou, S., & Ingram, L. O. (2000). Synergistic hydrolysis of carboxymethyl cellulose and acid-swollen cellulose by two endoglucanases (celz and cely) from *fromerwinia chrysanthemi*. *Journal of bacteriology*, 182(20), 5676-5682.