

No Considerable Evidence In Cytokines Profile Among Latent Toxoplasmosis Infertile Women. Cytokines In Toxoplasmosis Women

Alaa Tareq Shakir al-hassnawi^{#1}, Luma Hakim Ali² & Hassan Raheem Khudhur³

^{#1}*Department of Biology, College of Science -- University of Babylon, Iraq.*

²*Department of Biology, College of Science -- University of Kufa, Iraq.*

³*Department of Biology, College of Science -- University of Al-Qadisiya, Iraq*

^{#1}*alaatark79@yahoo.com*

²*h_luma@yahoo.com*

Abstract: Toxoplasma gondii is an obligate intracellular protozoan parasite causes toxoplasmosis, is one of the most prevalent human diseases in many countries [2, 19]. Reports on the prevalence of T. gondii infection among infertile females are limited, No previous reports about the relation between T. gondii infection and female infertility although about 21% of abnormal embryos and 24.2% of miscarriages and stillbirths have been related to T. gondii infection [21]. The present study aimed to survey the infertile females and Toxoplasma infection investigating a possible relationship with infertility. Sera from 18 Toxoplasma infected women and sera from 30 free Toxoplasma infertile women were tested by ELISA IgM and IgG toxoplasma antibodies, and the level of serum IL4, IL-8, IL10 and IL33 was tested using ELISA technique.

INTRODUCTION

Regardless of its function, *Toxoplasma gondii* is one of the obligatory intracellular protozoans,, which is responsible for common parasitic infections around the world [1], Toxoplasmosis is caused by infection with the *Toxoplasma gondii*, this type of infections produces a variety of clinical syndromes both in humans and animals, the main infection during pregnancy can result in disease transmission through the placenta and lead to hazardous consequences such as abortion, stillbirth, different degrees of mental or physical retardation, hydrocephalus, and blindness [2]. The infection stimulates cell mediated immunity and humoral immune response as antibody production, in addition to the role of cytokines, which essential for the host and control of intracellular infection [3].

Several serological methods have detected the immunoglobulin (IgG and IgM) antibodies against *T. gondii* in the serum and among the assays, ELISA shows high sensitivity and specificity [4]. IL8 has an important role in the innate immune response. It's often associated with inflammation. It has been cited as a pro-inflammatory mediator in Toxoplasmosis [5]. It is well recognized that T cell-mediated immunity plays a central role in the host response to intracellular pathogens [6]. T cell- mediated immunity and activated macrophages have been shown to play important roles in resistance to T cell-mediated immunity *T. gondii* infection [7].

The anti-inflammatory cytokine IL-10 plays an important role in reducing harmful pathological effects of inflammatory responses in *T.gondii* infection. IL-10 is a cytokine produced by DCs, macrophages, B-cells, Th2 cells and T-regulatory cells (Levings et al., 2002). IL-10-deficient mice showed elevated IL-12 levels and consequently increased IFN- γ

and TNF- α responses and intense hepatic inflammation and tissue necrosis [8]. During acute toxoplasmosis, IL-10 serves a dual role in the suppression of the host's cellular immune response.

IL-33 is a member of the IL-1 cytokine family, in the nucleus is associated with chromatin, [9, 10] upon cell stress or death, biologically active IL-33 is released and truncated by proteolytic cleavage [11] it may have a dual role in different inflammatory conditions, depending on the specific immune mechanisms underlying disease pathogenesis [12], infections induces proinflammatory cytokine and chemokine responses, which are reduced in absence of IL-33R/ST2 signaling [13].

The humoral immune response to *T. gondii* is rapid and intense, and forms the basis for useful diagnostic tests for the various forms of the disease, the present study aimed to detection of IL4, IL8, IL10 and IL33 by diagnostic method (ELISA) in serum infected and non-infected infertile women by toxoplasmosis.

MATERIALS AND METHODS

Exclusions Criteria

All exclusion criteria, such as family history, chronic and genetic disease, drinking and smoking, have previous abortions were account to exclude women from control one. After an interviewer managed we used forty eight enrolled infertile women.

Blood Collection

Statistical analysis was done using SPSS (Social Science Statistical Package) version 20 in which we use mean and standard deviation as descriptive statistics and LSD (the least significant difference) analysis of variance (ANOVA) for comparison between groups. The P value was considered significant if below 0.05.

Detection of Toxoplasma gondii Infections

Enzyme linked immunofluorescence assay (ELIFA) technique were used to confirm serum anti-toxoplasma IgG antibodies, the manual procedure accomplished by manufacture Biomerieux Company (France).

Enzyme linked immunosorbent assay (ELISA)

All cytokine biomarkers test in this study (IL-10,IL-33,IL-8 and IL-4) checked by sandwich ELISA briefly, serum were added in pre-coated micro ELISA plate wells, then a biotinylated detection antibody for each specific cytokine added after one half hour of incubation. HRP detection enzyme then added after 3 washes. TMP Specific substrate were used after 5 washes, blue color terminated by stop solution (diluted H₂SO₄). Optical density measured by using ELISA reader and result calculated by comparing O.D of sample by O.D of stander.

Study protocol and ethics

Study protocol approved and ethical issues done by local committee in college of science/ Babylon University, already the committee depends on principles of declaration Helsinki.

Statistical Analysis

All statistical analyses were performed according statistical software program (SPSS 10 Inc., Chicago, USA). All statistical comparisons were done by T-test. Variations were considered significant when P-value \leq 0.05

RESULTS

In the present study the enrolled women (18 *Toxoplasma* infected and 30 free *Toxoplasma* infertile women) were checked by ELIFA techniques to confirm the infection of *Toxoplasma gondii* (positive just for IgG antibodies). The percentage of infection in women was 37%. Seroprevalence of cytokines profile in *Toxoplasma* free and infected women showed no significant variation in interleukin-8, 33 and interleukin-10 concentration (Figures 1, 2 and 3). Our finding showed a significant variation in serum Interlukin-4, were infected infertile women showed higher level as compare with free infected one($P = 0.018$, Figure 4). In additions, personal correlations between anti-toxoplasma IgG antibodies and all serum cytokine profile showed no significant variation see figures 5,6,7,8 were $P > 0.050$.

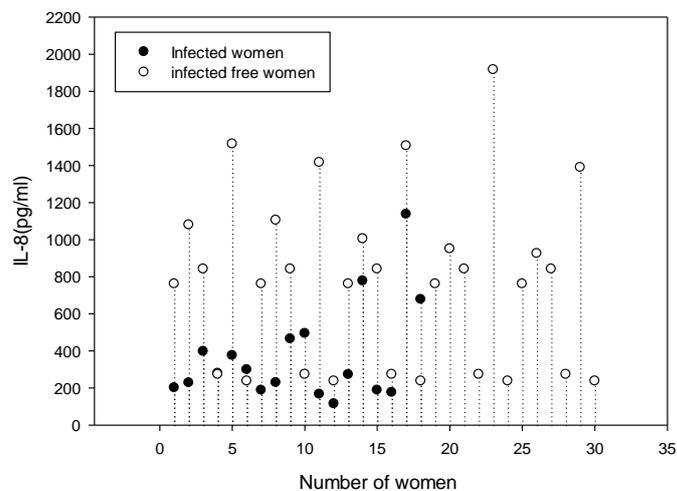


Figure 1. Comparison of mean concentration serum IL-8 Interleukin between patients and control Group.

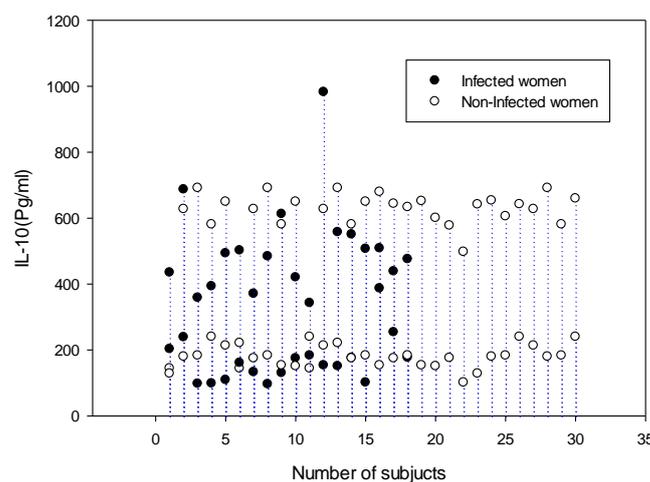


Figure 2. Comparison of mean concentration serum IL-10 Interleukin between patients and control Group.

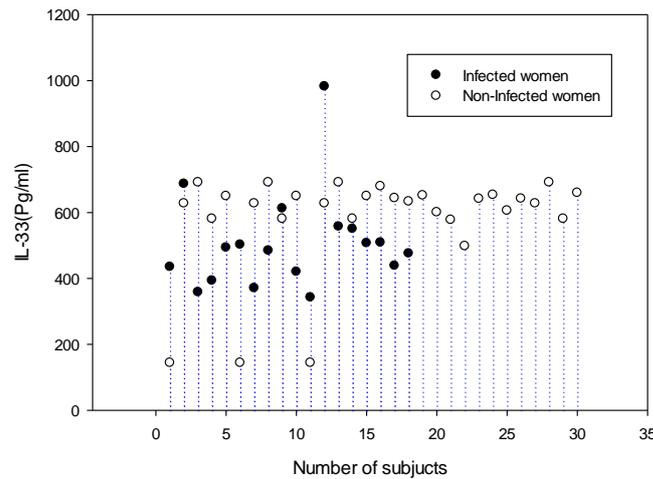


Figure 3. Comparison of mean concentration serum IL-33 Interleukin between patients and control Group.

The mean IL-4 were significantly higher in women patients as compared to control groups, (435.973+307.291pg/ml) versus (291.433+80.203pg/ml) respectively (p=0.0179). Figure (4).

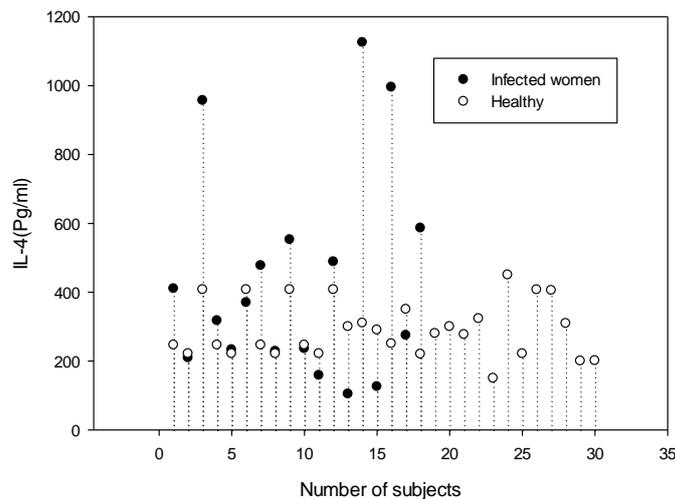


Figure 4. Comparison of mean concentration serum IL-4 Interleukin between patients and control Group

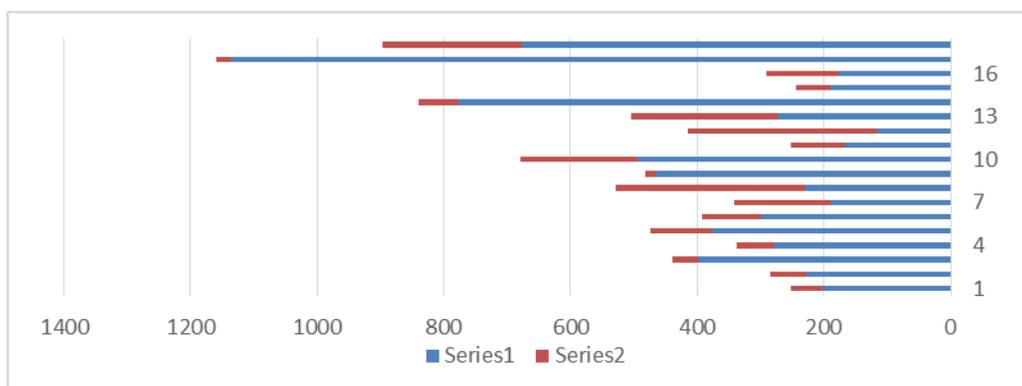


Figure 5. Serum IL-8 level among different levels of anti-Toxoplasma antibody (series 1=IL-8 series2= Anti-toxoplasma IgG antibodies). P-value= 0.101

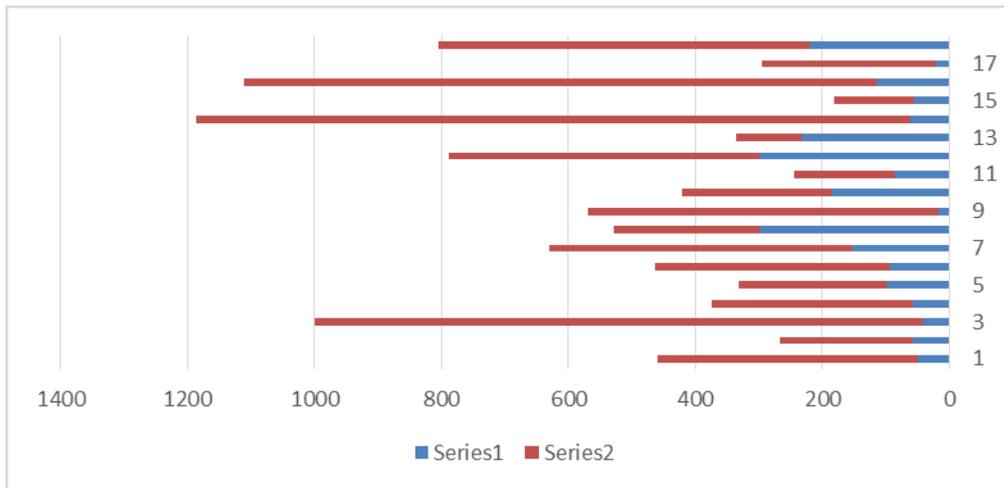


Figure 6. Serum IL-4 level among different levels of anti-Toxoplasma antibody (series 1=IL-4 series2= Anti-toxoplasma IgG antibodies). P-value= 0.797

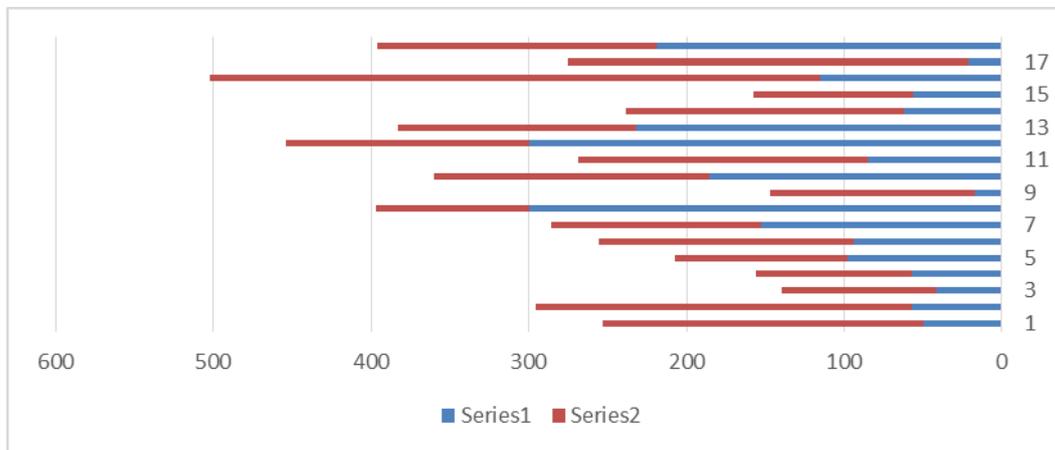


Figure 7. Serum IL-10 level among different levels of anti-Toxoplasma antibody (series 1=IL-10 series2= Anti-toxoplasma IgG antibodies). P-value= 0.742

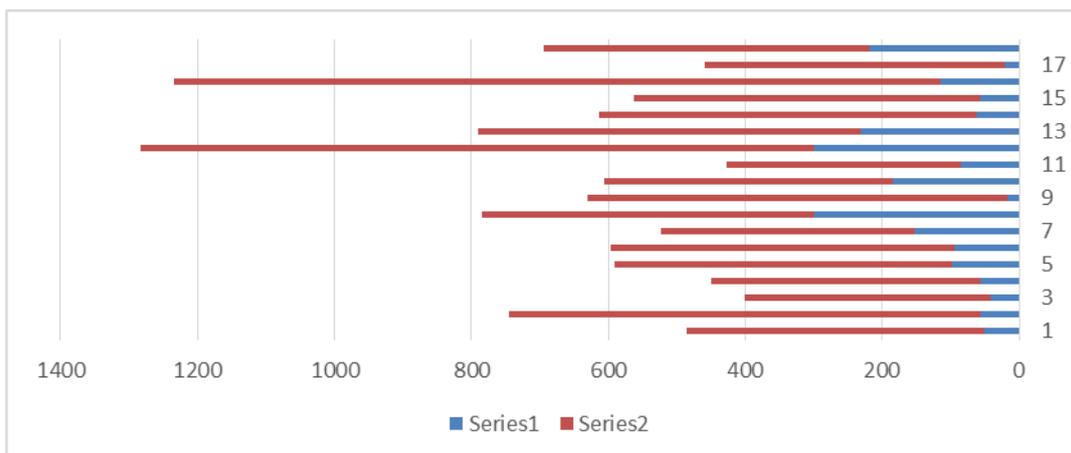


Figure 8. Serum IL-33 level among different levels of anti-Toxoplasma antibody (series 1=IL-33 series2= Anti-toxoplasma IgG antibodies). P-value= 0.497

DISCUSSION

It is known that toxoplasmosis has some unfavorable effects on the reproductive capacity of both men and women [14].

The present study is the first in Iraq, which showed there is no an association between toxoplasmosis and infertility women cytokines IL8, IL10 and IL33 concentration and showed no significant variation in the serum of infertility women infected with toxoplasmosis in comparison with infertility women uninfected (healthy control)

The statistical results indicated no significant variation of IL-8, IL-10 and IL-33 in infertile women patients' comparison with infertile women control, and may be due to the parasite alter or fail to stimulate secretion of the pro-inflammatory chemokines. The decrease level of IL-10 in patients in comparison with healthy control (Table 1) may be due to fail ability of the parasite to enhance TH2 cytokines among these was IL-10. However, IL-10 is strong enemy to macrophages capability in order to kill bacteria inside the cells microbes as well, examples are infections and *T. gondii* via numbers of pathogens, the presence of *T. gondii*, will lead to increasing in the IL-10 expression [15].

This results disagree with previous study done by El-Tantawy et al. [16] they found an association between toxoplasmosis and infertility women with significant higher prevalence ($p < 0.01$) of *T. gondii* infection in infertile female patients (61.85%) in Dakhalia governorate, Egypt in comparison with the pregnant women control group. Previous study in Iraq (2017) found that Interleukins IL-8 and IL-10 plays an important role in the resolution of *Toxoplasma gondii* infection, their concentration in women patients' serum of all age groups were increased in comparison to that observed in control groups [17].

The statistical results indicated increase of IL-4 in patients in comparison with healthy control of all ages (Figure 4). This increase is due to *T. gondii* stimulate secretion of the pro-inflammatory chemokines like IL-4. The chemokine response is dependent on invasion by live tachyzoites and subsequent host cell lysis. IL-4 is responsible for activation and recirculation of neutrophils and neutrophils can phagocyte and kill or inhibit tachyzoites of *Toxoplasma* and showed that human intestinal epithelial cells infected with *T. gondii* elicit rapid secretion of IL-4 [18].

CONCLUSION

The present study confirms that in SLE patients the antibodies to the antinucleosome are common. We have also revealed that mean antinucleosome antibodies are high in SLE groups in compared to healthy subjects or in rheumatoid arthritis patients. So, it might be a helpful addition to the laboratory tests that can aid with SLE diagnostics.

RECOMMENDATIONS

Our results found there is no significant variation in cytokines IL8 ,IL10 and IL33 of infertility women serum infected with toxoplasmosis in comparison with infertility women serum uninfected (healthy control)

REFERENCES

- [1] Mahmood, S.H., Hassani H.H. and Zghair K.H. (2010). Detection of B1 gene of *Toxoplasma gondii* in blood of pregnant and abortive women infected with this parasite. Iraqi Journal of Medical Sciences.
- [2] Elsheikha, H. M. (2008). Congenital toxoplasmosis: priorities for further health promotion action," Public Health, V. 122(4) : 335–353.

- [3] AL-Fertosi, R.B and Juma, A.S.M .(2006).Possible cellular expression IFN- γ in women with abortion infected with *Toxoplasma gondii*. Medical Journal of Islamic World Academy of Science. 16:121-34.
- [4] Saki, J.; Mohammadpour, M.; Moramezi, F. and Khademvatan, K. (2015). Seroprevalence of *Toxoplasma gondii* in Women Who Have Aborted in Comparison with the Women with Normal Delivery in Ahvaz, Southwest of Iran. Scientific World Journal. 10.1155.764369 :4.
- [5] Tenter A.M. and Heckerroth A.R. and Weiss LM. (2000). *Toxoplasma gondii*: from animals to human. Int. J Parasitol. 30:1217–58.
- [6] Šárka, K. and Jaroslav F. and Longer, S. M. .(2007). pregnancy and lower fetal development in women with latent "asymptomatic" toxoplasmosis. BMC. Infect Dis. 4:114.
- [7] Bliss, S.K.; Gavrilescu, L.C.; Alcaraz, A.; Denkers, E.Y. (2001).Neutrophil depletion during *Toxoplasma gondii* infection leads to impaired immunity and lethal systemic pathology. Infect Immun. 69:4898–905.
- [8] Gazzinelli, R.T.; Wysocka, M.; Hieny, S.; Scharon-Kersten T.; Cheever A., Kuhn R. In the absence of endogenous IL-10 mice acutely infected with *Toxoplasma gondii* succumb to a lethal immune response dependent on CD4+ Tcell and accompanied by over production of IL-12, INF-gamma TNF-alpha .(1996). J. Immunol. 157:798.
- [9] Schmitz, J.; Owyang, A.; Oldham, E.; Song, Y.; Murphy, E. and McClanahan, TK. (2005) .IL-33, an interleukin-1-like cytokine that signals via the IL-1 receptor-related protein ST2 and induces T helper type 2-associated cytokines. Immunity. 23:479–90..
- [10] Carrière, V.; Arshad, M.I.; Le Seyec, J.; Lefevre, B.; Farooq, M.; Jan, A. (2017).Endogenous IL-33 deficiency exacerbates liver injury and increases hepatic influx of neutrophils in acute murine viral hepatitis. Mediators of Inflammation. 1359064:15.
- [11] Cayrol, C.; Duval, A.; Schmitt, P.; Roga, S.; Camus, M. and Stella, A. (2018). Environmental allergens induce allergic inflammation through proteolytic maturation of IL-33. Nation. Immunol. 19:375–85.
- [12] Liew, F.Y.; Girard, J.P; Turnquist, H.R.(2016). Interleukin-33 in health and disease. Nat Rev Immunol. 16(11):676–689.
- [13] Ryffel, B.; Huang, F.; Robinet, P.; Panek, C.; Couillin, I.;Erard, F.; Piotet, J.; Le Bert, M.; Mackowiak, C.; Arias, M.T.; Dimier-Poisson, I. and Zheng S.G. .(2019).Blockade of IL-33R/ST2 Signaling Attenuates *Toxoplasma gondii* Ileitis Depending on IL-22 Expression . Front immunol. V.10 :702.
- [14] Sarkar MD, Anuradha B, Sharma N, Roy RN.(2012). Seropositivity of toxoplasmosis in antenatal women with bad obstetric history in a tertiary-care hospital of Andhra Pradesh, India. J Health Popul Nutr, 30 (1): 87-92.
- [15] Gazzinelli, R.T.; Oswald, I.P.; Sher, A. IL-10 inhibits parasite killing and nitrogen oxide production by IFN-gammaactivated macrophages.(1992). The Journal of Immunology. V. 148(6) : 1792-6.
- [16] El-Tantawy, N.; Taman, A. and Shalaby, H.(2014). Toxoplasmosis and Female Infertility: Is there a Co-Relation. American Journal of Epidemiology and Infectious Disease, Vol. 2, No. 1, 29-32.
- [17] Mohamed, K. I. A.; Khadhum, M. S.;Abu-Al-Ess, H. Q. M.;Ali, S. H. M.;Al-Fukhar, S. A.;Al-Wattar, W. M. A.;Hamoudi, S. R. and Mousa, J. M. (2017) . The Effect of *Toxoplasma gondii* on Interleukin-8, Interleukin-10, Leukotriene B4 and Calcium Levels in Aborted Women. International Journal of Medical Research & Health Sciences. 6(11): 76-82.

- [18] Ju, C.H.; Chockalipgam,A. and Leifer, C.A. (2009).Early response of mucosal epithelial cells during *Toxoplasma gondii* infection.” *The Journal of Immunology*.V. 183(11) : 7420-27.
- [19] Jones J. L., Kruszon-Moran D., Wilson M., McQuillan G., Navin T., McAuley J. B. .(2001).*Toxoplasma gondii* infection in the United States: seroprevalence and risk factors. *American Journal of Epidemiology*. 154(4):357–365.
- [20] Locksley, Richard M., John Fankhauser, and William R. Henderson. “Alteration of leukotriene release by macrophages ingesting *Toxoplasma gondii*.(1985). *Proceedings of the National Academy of Sciences V. 82(20) : 6922-26*.
- [21] Nowakowska D, Respondek-Liberska M, Gola E, Stray Pedersen B, Szaflik K, Dzbenski T, Wilczynski J. (2005) . Too late prenatal diagnosis of fetal toxoplasmosis: a case report. *Fetal diagnosis and therapy*, 20, 190-193.