In Vitro Evaluation Of Trichoderma Species Against Colletotrichum Falcatum Causing Red Rot Of Sugarcane

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Abstract: Red rot caused by Colletotrchum falcatum is one of the most important diseases of Sugarcane. In the present investigation, inhibitory effects of different Trichoderma species were observed in lab condition. It was found that all five Trichoderma species were efficient in mycelia growth inhibition of Colletotrichum falcatum. Trichoderma harzianum gave best result in dual culture with growth inhibition of 84.4% against whereas T. koningii showed minimum growth inhibition of 76.2%.

Keywords: Colletotrichum, Trichoderma, inhibition

1. INTRODUCTION

Soil biodiversity plays a very important role in the sustainability of agriculture systems and it also indicates the level of health of soil, particularly while considering the richness of microorganisms which are involved in plant disease control. The use of microbial diversity to manage disease of crop plants falls into the category of biological control. Researchers are therefore attempting to enhance the effectiveness of antagonists in the cropping field, thus increasing suppressive ness. Biocontrol of plant diseases by other microorganisms is found to be a more effective and environmentally beneficial substitute to the harmful chemical fungicides (Shalini and Kotasthane, 2007, ChitraMani & Kumar, P. (2020); Sharma, M., & Kumar, P. (2020); Chand, J., & Kumar, P. (2020); Naik, M., & Kumar, P. (2020); Kumar, P., & Naik, M. (2020); Kumar, P., & Dwivedi, P. (2020); Devi, P., & Kumar, P. (2020); Kumari, P., & Kumar, P. (2020); Kaur, S., & Kumar, P. (2020); Devi, P., & Kumar, P. (2020); Sharma, K., & Kumar, P. (2020); Kumar, S. B. P. (2020); Devi, P., & Kumar, P. (2020); Chand, J., & Kumar, P. (2020). Trichoderma species are best known to contain antifungal activity against plant diseases since 1930s (Hjeljord and Tronsmo, 1998). Their different species and isolates are available commercially (Freeman et al., 2004). T. harzianum solely or in mixture with different Trichoderma species or adjuvant, has been useful in management of several diseases like damping-off in radish, maize and soybean (Lifshitz et al., 1985) and tomato grey mould disease (Migheli et al., 1994). Sugarcane is one of the important commercial crops playing major role in agriculture and industrial economy of the country.

2. MATERIAL AND METHODS

Five different species presented in **Table 1** were assessed for comparative efficacy against Collectrichum falcatum by dual culture method. Nine mm diameter disc of test fungus and the antagonistic fungi were cut and were kept opposite to each other at a distance of 5 mm

from the edge of Petri dish. Same disc of test fungus was placed on another petri plate containing PDA, which served as Untreated (Control). Individual treatment was duplicated 3 times and incubated at 25 ± 2^{0} C. Per cent growth inhibition of test pathogen was found out by using given below formula (Vincent, 1947 and Behzad et al., 2008).

Inhibition % =
$$\frac{\text{CONTROL} - \text{TREATED}}{\text{CONTROL}} \times 100$$

Where, C = Mycelial growth of the test Pathogen in controlled plate (mm) T = Mycelial growth of the test Pathogen in treatment (mm)

Tuble 1. Else of Thenodelina species used against concentration falcatain		
Treatment	Name of Bio agent /Botanical	
T1	Trichoderma viride	
T2	Trichoderma hamatum	
T3	Trichoderma harzianum	
T4	Trichoderma koningii	
T5	Trichoderma virens	

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3. RESULT AND DISCUSSION

Effect of five different species (Table 2) were evaluated under in vitro condition. Trichoderma isolates were cultured along with Colletotrichum falcatum in petriplates for a week and it was found that all the species of Trichoderma reduced themycelial growth of Colletotrichum falcatum. Trichoderma harzianum gave best result in dual culture with minimum radial growth of 14.5mm and maximum growth inhibition of 84.4% followed by T.hamatum with radial growth of 15mm and growth inhibition of 83.3mm, T.viride with 82.2mm growth inhibition, T.virens with 77.2% growth inhibition whereas T. koningii showed minimunm growth inhibition of 76.2% with maximum mycelial growth of 21.5mm. Singh et al. (2004) observed inhibitory activity of different Trichoderma isolates against Colletotrichum falcatum. Webster and Lomas (1964) reported that Trichoderma viride produces gliotoxin and viridin which easily inhibit the growth of pathogens (Kumar, P. (2019); Kumar, D., Rameshwar, S. D., & Kumar, P. (2019); Dev, S. R., & Kumar, P. (2019); Kumar et al. (2019); Dey, S. R., & Kumar, P. (2019); Kumar, P., & Pathak, S. (2018); Kumar, P., & Dwivedi, P. (2018); Kumar, P., & Pathak, S. (2018); Kumar et al., 2018; Kumar, P., & Hemantaranjan, A. (2017); Dwivedi, P., & Prasann, K. (2016). Kumar, P. (2014); Kumar, P. (2013); Kumar et al. (2013); Prasann, K. (2012); Kumar et al. (2011); Kumar et al. (2014).

Table2. Effect of different species of Trichoderma on the growth of Collectotrichum falcatum

S.No.	Trichoderma species	Colony	Percent growth
		diameter(mm)	inhibition (%)
1.	T.harzianum	14.5	84.4
2.	T.koningii	21.5	76.2
3.	T.hamatum	15	83.3
4.	T.viride	16	82.2
5.	T.virens	20.5	77.2

Control	90	00
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REFERENCES

- Freeman, S., Minz, D., Kolesnik, I., Barbul, O., Zveibil, A., Maymon, M., ... & Dag, A. (2004). Trichoderma biocontrol of Collectorichum acutatum and Botrytis cinerea and survival in strawberry. European Journal of Plant Pathology, 110(4), 361-370.
- [2] Hajieghrari, B., Torabi-Giglou, M., Mohammadi, M. R., & Davari, M. (2008). Biological potantial of some Iranian Trichoderma isolates in the control of soil borne plant pathogenic fungi. African Journal of Biotechnology, 7(8).
- [3] Harman, G. E., Björkman, T., Ondik, K., & Shoresh, M. (2008). Changing paradigms on the mode of action and uses of Trichoderma spp. for biocontrol. Outlooks on pest management, 19(1), 24.
- [4] Hjeljord, L., & Tronsmo, A. (1998). Trichoderma and Gliocladium. biological control: an overview. In: Trichoderma & Gliocladium: Enzymes, biological control and commercial applications. Harman GE, Kubice CP.(Eds), 2, 131-151.
- [5] Lifshitz R, Windham M T and Baker R 1986 Mechanism of biological control of preemergence damping-off of pea by seed treatment with Trichoderma spp. Phytopathology 76, 720–725.
- [6] Migheli, Q., Herrera-Estrella, A., Avataneo, M., & Gullino, M. L. (1994). Fate of transformed Trichoderma harzianum in the phylloplane of tomato plants. Molecular ecology, 3(2), 153-159.
- [7] Shalini, S., & Kotasthane, A. S. (2007). Parasitism of Rhizoctonia solani by strains of Trichoderma spp. EJEAF Chemistry, 6, 2272-2281.
- [8] Singh, V., Lal, R. J., Lal, S., Sinha, O. K., Singh, A. P., & Srivastava, S. N. (2004, September). Bioefficient strains of Trichoderma harzianum against red rot pathogen of sugarcane. In Proceedings of Symposium on Recent Advances in Fungal Bioagents and Their Social Benefit, Zonal Chapter, Ann. Meet., Mycol. & Pl. Pathol, NBRI, Lucknow (pp. 47-48).
- [9] Vincent, J. M. (1947). Distortion of fungal hyphae in the presence of certain inhibitors. Nature, 159(4051), 850-850.
- [10] ChitraMani, P. K. (2020). Evaluation of antimony induced biochemical shift in mustard. Plant Archives, 20(2), 3493-3498.
- [11] Sharma, M., & Kumar, P. (2020). Biochemical alteration of mustard grown under tin contaminated soil. Plant Archives, 20(2), 3487-3492.
- [12] Chand, J., & Kumar, P. (2020). Yield attribute shift of mustard grown under cadmium contaminated soil. Plant Archives, 20(2), 3518-3523.
- [13] Naik, M., & Kumar, P. (2020). Role of growth regulators and microbes for metal detoxification in plants and soil. Plant Archives, 20(2), 2820-2824.
- [14] Kumar, P., & Naik, M. (2020). Biotic symbiosis and plant growth regulators as a strategy against cadmium and lead stress in chickpea. Plant Archives, 20(2), 2495-2500.
- [15] Kumar, P., & Dwivedi, P. (2020). Lignin estimation in sorghum leaves grown under hazardous waste site. Plant Archives, 20(2), 2558-2561.
- [16] Devi, P., & Kumar, P. (2020). Concept and Application of Phytoremediation in the Fight of Heavy Metal Toxicity. Journal of Pharmaceutical Sciences and Research, 12(6), 795-804.

- [17] Kumari, P., & Kumar, P. (2020). Trichoderma fungus in mitigation of rhizosphere arsenic: with special reference to biochemical changes. Plant Archives, 20(2), 3512-3517.
- [18] Kaur, S., & Kumar, P. (2020). Ameliorative effect of trichoderma, rhizobium and mycorrhiza on internodal length, leaf area and total soluble protein in mung bean (Vigna radiata [L.] R. Wilazek) under drought stress. Journal of Pharmacognosy and Phytochemistry, 9(4), 971-977.
- [19] Devi, P., & Kumar, P. (2020). Effect of bioremediation on internodal length and leaf area of maize plant cultivated in contaminated soil with chromium metal. Journal of Pharmacognosy and Phytochemistry, 9(4), 1408-1413.
- [20] Sharma, K., & Kumar, P. (2020). Mitigating the effect of biofertilizers on morphological and biochemical level in pearl millet grown under mercury toxicity. Journal of Pharmacognosy and Phytochemistry, 9(4), 955-961.
- [21] Kumar, S. B. P. (2020). Salinity stress, its physiological response and mitigating effects of microbial bio inoculants and organic compounds. Journal of Pharmacognosy and Phytochemistry, 9(4), 1397-1303.
- [22] Devi, P., & Kumar, P. (2020). Enhancement effect of biofertilizers on germination percentage and plant height in maize grown under chromium toxic soil. Journal of Pharmacognosy and Phytochemistry, 9(4), 702-707.
- [23] Chand, J., & Kumar, P. (2020). Biochemical shift of mustard grown under cadmium contaminated soil. Journal of Pharmacognosy and Phytochemistry, 9(3), 178-183.
- [24] Kumar, P. (2019). Evaluation Of Internodal Length And Node Number Of Pea Treated With Heavy Metal, Polyamines And Glomus. Journal of the Gujarat Research Society, 21(10s), 518-523.
- [25] Kumar, D., Rameshwar, S. D., & Kumar, P. (2019). Effect Of Intergated Application Of Inorganic And Organic Fertilizers On The Roots Of Chickpea. Plant Archives, 19(1), 857-860.
- [26] Dey, S. R., & Kumar, P. (2019). Analysis of Available Nitrogen of Wheat Cultivated Soil Treated with Organic and Inorganic Source of Fertilizers. Int. J. Curr. Microbiol. App. Sci, 8(8), 2986-2990.
- [27] Kumar, P., Siddique, A., Thakur, V., & Singh, M. (2019). Effect of putrescine and glomus on total reducing sugar in cadmium treated sorghum crop. Journal of Pharmacognosy and Phytochemistry, 8(2), 313-316.
- [28] Dey, S. R., & Kumar, P. (2019). Cadmium induced biochemical shift in maize. Journal of Pharmacognosy and Phytochemistry, 8(1), 2038-2045.
- [29] Kumar, P., & Pathak, S. (2018). Short-Term Response of Plants Grown under Heavy Metal Toxicity. Heavy Metals, 69.
- [30] Kumar, P., & Dwivedi, P. (2018). Plant lectins, agricultural advancements and mammalian toxicity. Molecular Physiology of Abiotic Stresses in Plant Productivity, 360.
- [31] Kumar, P., & Pathak, S. (2018). Nitric oxide: a key driver of signaling in plants. MOJ Eco Environ Sci, 3(3), 145-148.
- [32] Kumar, P., Pathak, S., Amarnath, K. S., Teja, P. V. B., Dileep, B., Kumar, K., ... & Siddique, A. (2018). Effect of growth regulator on morpho-physiological attributes of chilli: a case study. Plant Archives, 18(2), 1771-1776.
- [33] Kumar, P., & Hemantaranjan, A. (2017). Iodine: a unique element with special reference to soil-plant-air system. Advances in Plant Physiology (Vol. 17), 314.
- [34] Dwivedi, P., & Prasann, K. (2016). Objective plant physiology. Objective plant physiology., (Ed. 2).

- [35] Kumar, P. (2014). Significance of soil-root system and aquaporins for water homeostasis in plant-a review. Advances in Plant Physiology (Vol. 15), 15, 324.
- [36] Kumar, P. (2013). Food Security and Nutritional Safety: A Challenge Ahead. Journal of Functional and Environmental Botany, 3(1), 12-19.
- [37] Prasann, K., Biswapati, M., & Padmanabh, D. (2013). Combating heavy metal toxicity from hazardous waste sites by harnessing scavenging activity of some vegetable plants. Vegetos, 26(2), 416-425.
- [38] Prasann, K. (2012). Feeding the future: crop protection today. Acta Chimica and Pharmaceutica Indica, 2(4), 231-236.
- [39] Kumar, P., & Dwivedi, P. (2011). Future Habitat Loss: Greatest Threat to the Soil Microbial Biodiversity. Journal of Functional And Environmental Botany, 1(2), 82-90.
- [40] Kumar, P., Singh, B. N., & Dwivedi, P. Plant Growth Regulators, Plant Adaptability And Plant Productivity: Areview On Abscisic Acid (Aba) Signaling In Plants Under Emerging Environmental Stresses. Sustaining Future Food Security In Changing Environments, 81.